

Novel peptides with increased + charge and
hydrophobicity by substituting one or more
amino acids of CA-MA peptide and pharmaceutical
compositions containing thereof

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FIELD OF THE INVENTION

The present invention relates to novel peptides with increased + charge and hydrophobicity by substituting one or more amino acids of CA-MA peptide in which cecropin A (CA) and magainin 2 (MA) were conjugated and pharmaceutical compositions containing thereof. More precisely, the present invention relates to synthetic peptides prepared by substituting one or more amino acids of CA-MA peptide represented by the

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SEQ. ID. NO: 1 with amino acids having + charge and hydrophobicity and anti-bacterial, anti-fungal and anticancer compositions containing thereof. The synthetic peptides of the present invention have no cytotoxicity but have excellent anti-bacterial, anti-

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fungal and anticancer activity, leading in an effective use thereof as a safe anticancer agent and antibiotics.

BACKGROUND

Bacteria infection is one of the most common but

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fatal causes for human diseases. Infection has been successfully treated by antibiotics, but the abuse of

antibiotics brought another problem that bacteria now might have resistance against antibiotics. In fact, the speed which bacteria are adapting and having resistance against new antibiotics outruns that of developing new antibiotics analogues. For example, fatal *Enterococcus faecalis*, *Mycobacterium tuberculosis* and *Pseudomonas aeruginosa* are known to have raised their resistance against every possible antibiotics (Stuart B. Levy, *Scientific American*, 1998, 46-53).

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Tolerance is different from resistance against antibiotics, and it was firstly found in *Pneumococcus* sp. in 1970s, which provided an important clue for disclosing the mechanism of penicillin (Tomasz, et al., *Nature*, 1970, 227, 138-140). Some bacteria species having tolerance stopped growing under the ordinary concentration of antibiotics but never died. Tolerance is caused by that the activity of autolytic enzyme of bacteria, like autolysin, is suppressed when the antibiotics inhibit cell wall synthetase. Penicillin can kill bacteria by activating endogenous hydrolytic enzyme; on the other hand, bacteria can survive by restraining the activity thereof even when being treated with antibiotics.

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It is a clinical hot issue that bacteria are having tolerance against various antibiotics since

infection cannot be effectively cured with antibiotics due to the tolerance (Handwerger and Tomasz, *Rev. Infect. Dis.*, 1985, 7, 368-386). Again, once bacteria have tolerance, they can have resistance, which helps that bacteria survive under antibiotics treatment. Such bacteria can acquire new genetic elements having resistance against antibiotics, thus they can grow even under antibiotics treatment. Actually, bacteria having resistance have tolerance, too (Liu and Tomasz, *J. Infect. Dis.*, 1985, 152, 365-372). Thus, it is urgent to develop novel antibiotics, which can kill antibiotics-resistant bacteria.

There are two types of tolerance in the aspect of its mechanism. The first one is phenotypic tolerance, which occurs when the growing speed decreases in all kinds of bacteria (Tuomanen E., *Revs. Infect. Dis.*, 1986, 3, S279-S291), and the second one is genotypic tolerance acquired by mutation in a certain type of bacteria. For both cases, down regulation of autolysin activation is basically occurring. In the case of phenotypic tolerance acquired by outside stimulus, down regulation takes place temporally while down regulation occurs permanently in the case of genotypic tolerance acquired by mutation, which cause the change of hemolysis regulating routes. Autolysin deficiency is believed to cause the simplest genotypic tolerance, but

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second group is α -helical amphiphilic peptides, and the third group is proline-rich peptides (Mayasaki et al., *Int. J. Antimicrob. Agents*, 1998, 9, 269-280). Those anti-bacterial peptides are known to play an important role in host-defense and congenital immune system (Boman, H. G., *Cell*, 1991, 65, 205; Boman, H. G., *Annu. Rev. Microbiol.*, 1995, 13, 61). The anti-bacterial peptides have many different structures depending on amino acid sequences, and the most common structure is amphiphilic α -helical structure having no cysteine, just like cecropin, an anti-bacterial peptide found in insects.

Among those peptides, the anti-bacterial activity of amphiphilic peptides has been studied and the development of antibiotics using the amphiphilic peptides has been tried. As of today, magainin 2 (MA), cecropin A (CA) and melittin (ME) have been reported as amphiphilic peptides.

Amphiphilic peptides of cecropin group were first found in a fruit fly and later in a silkworm pupa and in a pig intestine, too. While cecropin A was reported to have high anti-bacterial activity but low anti-fungal and anticancer activity (Boman, H. G. and Hultmark, D., *Annu. Rev. Microbiol.*, 1987, 41, 103), magainin 2 was known not to have cytotoxicity but to have appreciable anti-bacterial, anti-fungal, anticancer and anti-protozoa activity (Zasloff, M.,

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SUMMARY OF THE INVENTION

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excellent anti-bacterial, anti-fungal and anticancer activity without cytotoxicity.

It is another object of this invention to provide pharmaceutical compositions for anti-bacterial, anti-fungal and anticancer agent containing the above synthetic peptides.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 is photographs showing the number of colonies on LB agar plate, in which *Bacillus subtilis* was treated with synthetic peptide of the present invention,

A: Positive control,

B: CA-MA peptide,

C: Synthetic peptide represented by the SEQ. ID. NO: 2

FIG. 2 is photographs showing the number of colonies on NB+0.5% NaCl agar plate, in which *Pseudomonas aeruginosa* was treated with synthetic peptide of the present invention,

A: Positive control,

B: CA-MA peptide,

C: Synthetic peptide represented by the SEQ. ID. NO: 2

FIG. 3 is SEM (scanning electron microscopy) microphotographs showing the result of treating synthetic peptide of the present invention to *Bacillus subtilis*,

- 5 A: Positive control,
 B: CA-MA peptide,
 C: Synthetic peptide represented by the SEQ. ID.
 NO: 2

10 FIG. 4 is SEM microphotographs showing the result of treating synthetic peptide of the present invention to *Pseudomonas aeruginosa*,

- A: Positive control,
 B: CA-MA peptide,
 15 C: Synthetic peptide represented by the SEQ. ID.
 NO: 2

FIG. 5 is graphs showing the dynamic condition of lipid membrane after treating synthetic peptide of the present invention to *Bacillus subtilis* and *Pseudomonas aeruginosa*,

- 20 A: Dynamic condition of lipid membrane of *Bacillus subtilis*,
 B: Dynamic condition of lipid membrane of *Pseudomonas aeruginosa*,
 25 ● : CA-MA peptide,
 □ : Synthetic peptide represented by the SEQ. ID.

novel peptides and their derivatives with increased + charge and hydrophobicity by substituting one or more amino acids of cecropin A and magainin 2 conjugated CA-MA peptide represented by the SEQ. ID. NO: 1.

5 Peptides and their derivatives of the present invention were synthesized to have increased + charge and hydrophobicity by substituting a few amino acids including hinge region of CA-MA peptide which was prepared by conjugating 1-8 amino acid region of
10 amphiphilic helical CA and 1-12 amino acid region of MA, and represented by the SEQ. ID. NO: 1, with other amino acids.

 In order to prepare synthetic peptides of the present invention, the present inventors used
15 Merrifield's liquid solid phase method in which Fmoc (9-fluorenylmethoxycarbonyl) was used as a protecting group (Merrifield, R. B., *J. Am. Chem. Soc.*, 1963, 85, 2149). Every synthetic peptide with increased + charge and hydrophobicity by substituting one or more amino
20 acids including hinge region of CA-MA peptide represented by the SEQ. ID. NO: 1 could be peptide of the present invention. Especially, peptides and their derivatives prepared by substituting glycine-isoleucine-glycine residing at hinge region of CA-MA
25 peptide represented by the SEQ. ID. NO: 1 with proline each, substituting 4th leucine, 8th isoleucine, 14th leucine, 15th histidine with lysine each, and

substituting 5th phenylalanine, 6th lysine, 12th lysine, 13th phenylalnine, 16th serine, 17th alanine, 20th phenylalanine with leucine were preferred.

5 The peptide synthesized as above was isolated and purified, after which the purity thereof was confirmed. As a result, the purity of the peptide was over 95%, and the molecular weight obtained by MALDI (Matrix-Assisted Laser Desorption Ionization) mass spectrometry
10 (Hill, et al., *Rapid Commun. Mass Spectrometry*, 1991, 5, 395) was the same as the molecular weight obtained by calculation of amino acids. Therefore, it was confirmed that the peptide having correct amino acid sequence represented by the SEQ. ID. NO: 2 was
15 synthesized.

 The present invention also provides anti-bacterial, anti-fungal and anticancer pharmaceutical compositions containing the above peptides and their derivatives.

 To confirm if the peptides and their derivatives
20 of the present invention can be used for anti-bacterial, anti-fungal and anticancer agents, the present inventors have measured the anti-bacterial activity of the synthetic peptides by measuring minimal inhibitory concentration (referred as "MIC" hereinafter).

25 Synthetic peptide of the present invention represented by the SEQ. ID. NO: 2 was used to measure MIC value to each strain. As a result, synthetic

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Considering all those results together, the

synthetic peptide of the present invention represented by the SEQ. ID. NO: 2 was confirmed to have excellent anti-bacterial, anti-fungal and anticancer activity without cytotoxicity, so that the peptide can be effectively used as a safe anti-bacterial, anti-fungal and anticancer treatment agent.

Peptides and their derivatives of the present invention can be administered orally or parenterally. The compounds of the present invention can be prepared for oral or parenteral administration by mixing with generally-used fillers, extenders, binders, wetting agents, disintegrating agents, diluents such as surfactant, or excipients. The present invention also includes pharmaceutical formulations in dosage units. This means that the formulations are presented in the form of individual parts, for example tablets, coated tablets, capsules, pills, suppositories and ampules, the active compound content of which corresponds to a fraction or a multiple of an individual dose. The dosage units can contain, for example, 1, 2, 3 or 4 individual doses or 1/2, 1/3 or 1/4 of an individual dose. An individual dose preferably contains certain amount of active compound, which is administered in one application and which usually corresponds to a whole, one half, one third, or a quarter of a daily dose. Non-toxic inert pharmaceutically suitable excipients

are to be understood as solid, semi-solid or liquid diluents, fillers and formulation auxiliaries of all types. Preferred pharmaceutical formulations which may be mentioned are tablets, coated tablets, capsules, pills, granules, suppositories, solutions, suspensions and emulsions, pastes, ointments, gels, creams, lotions, dusting powders and sprays. Solid formulations for oral administration are tablets, pill, dusting powders and capsules. Liquid formulations for oral administrations are suspensions, solutions, emulsions and syrups, and the abovementioned formulations can contain various excipients such as wetting agents, sweeteners, aromatics and preservatives in addition to generally-used simple diluents such as water and liquid paraffin. Tablets, coated tablets, capsules, pills and granules can contain the active compound or compounds in addition to the customary excipients, such as (a) fillers and extenders, for example starches, lactose, sucrose, glucose, mannitol and silicic acid, (b) binders, for example carboxymethylcellulose, alginates, gelatine and polyvinylpyrrolidone, (c) humectants, for example glycerol, (d) disintegrating agents, for example agar-agar, calcium carbonate and sodium carbonate, (e) solution retarders, for example paraffin, and (f) absorption accelerators, for example quaternary ammonium compounds, (g) wetting agents, for example cetyl alcohol and glycerol monostearate, (h)

adsorbents, for example kaolin and bentonite, and (i) lubricants, for example talc, calcium stearate, magnesium stearate, and solid polyethylene glycols, or mixtures of the substances listed under (a) to (i).

5 The tablets, coated tablets, capsules, pills and granules can be provided with the customary coatings and shells, optionally containing opacifying agents, and can also be of a composition such that they release the active compound or compounds only or preferentially

10 in a certain part of the intestinal tract, if appropriate in a delayed manner, examples of embedding compositions which can be used would be polymeric substances and waxes. If appropriate, the active compound or compounds can also be presented in

15 microencapsulated form with one or more of the abovementioned excipients. Formulations for parenteral administration are sterilized aqueous solutions, water-insoluble excipients, suspensions, emulsions, and suppositories. Suppositories can contain, in addition

20 to the active compound or compounds, the customary water-soluble or water-insoluble excipients, for example polyethylene glycols, fats, for example cacao fat, and higher esters (for example C14-alcohol with C16-fatty acid) or mixtures of these substances.

25 Ointments, pastes, creams and gels can contain, in addition to the active compound or compounds, the customary excipients, for example animal and vegetable

fats, waxes, paraffins, starch, tragacanth, cellulose derivatives, polyethylene glycols, silicones, bentonites, silicic acid, talc and zinc oxide, or mixtures of these substances. Dusting powders and
5 sprays can contain, in addition to the active compound or compounds, the customary excipients, for example lactose, talc, silicic acid, aluminum hydroxide, calcium silicate and polyamide powder, or mixtures of these substances. Sprays can additionally contain the
10 customary propellants, for example chlorofluorohydrocarbons. Solutions and emulsions can contain, in addition to the active compound or compounds, the customary excipients, such as solvents, solubilizing agents and emulsifiers, for example
15 water, ethyl alcohol, isopropyl alcohol, ethylcarbonate, ethyl acetate, benzyl alcohol, benzyl benzoate, propylene glycol, 1,3-butylene glycol, dimethylformamide, oils, in particular cottonseed oil, groundnut oil, corn germ oil, olive oil, castor oil and
20 sesame oil, glycerol, glycerol formal, tetrahydrofurfuryl alcohol, polyethylene glycols and fatty acid esters of sorbitan, or mixtures of these substances. For parenteral administration, the solutions and emulsions are also be in a sterile form
25 which is isotonic with blood. Suspensions can contain, in addition to the active compound or compounds, the customary excipients, such as liquid diluents, for

example water, ethyl alcohol and propylene glycol, and suspending agents, for example ethoxylated isostearyl alcohols, polyoxyethylene sorbitol and sorbitan esters, microcrystalline cellulose, aluminum metahydroxide, bentonite, agar-agar and tragacanth, or mixtures of these substances. The formulation forms mentioned can also contain coloring agents, preservatives and additives that improve the smell and taste, for example peppermint oil and eucalyptus oil, and sweeteners, for example saccharin. The abovementioned pharmaceutical formulations can also contain other pharmaceutical active compounds in addition to the compounds according to the present invention. The abovementioned pharmaceutical formulations are prepared in the customary manner by known methods, for example by mixing the active compound or compounds with the excipient or excipients.

The therapeutically active compounds should preferably be present in the abovementioned pharmaceutical formulations in a concentration of about 0.1 to 99.5, preferably about 0.5 to 95% by weight of the total mixture.

The formulations mentioned can be used on humans and animals orally, rectally, parenterally (intravenously, intramuscularly or subcutaneously), intracisternally, intravaginally, intraperitoneally or locally (dusting powder, ointment, drops) and for the

therapy of infections in hollow spaces and body cavities. Possible suitable formulations are injection solutions, solutions and suspensions for oral therapy and gels, infusion formulations, emulsions, ointments or drops, ophthalmological and dermatological formulations, silver salts and other salts, eardrops, eye ointments, dusting powders or solutions can be used for local therapy. In the case of animals, intake can also be in suitable formulations via the feed or drinking water. Gels, powders, dusting powders, tablets, delayed release tablets, premixes, concentrates, granules, pellets, boli, capsules, aerosols, sprays and inhalants can furthermore be used on humans and animals. The compounds according to the present invention can moreover be incorporated into other carrier materials, such as for example, plastics (chain of plastic for local therapy), collagen or bone cement.

In general, it has proved advantageous both in human and in veterinary medicine to administer the active compound or compounds according to the present invention in total amounts of about 0.1 to about 2 mg/kg, preferably 0.5 to 1 mg/kg of body weight, 1-3 times every 24 hours, if appropriate in the form of several individual doses, to achieve the desired results. However, it may be necessary to deviate from the dosages mentioned, and in particular to do so as a

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EXAMPLES

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Example 1: Synthesis of oligonucleotide represented by
the SEQ. ID. NO: 1

In order to synthesize the peptide of the present invention represented by the SEQ. ID. NO: 2, we, the present inventors used Merrifield's liquid solid phase method in which Fmoc (9-fluorenylmethoxycarbonyl) was used as a protecting group (Merrifield, R. B., *J. Am. Chem. Soc.*, 1963, 85, 2149). For the peptide having -NH₂ type carboxyl terminal, rink amide MBHA-resin was used as a starting material. And, Fmoc-amino acid-Wang resin (SynPep Corporation) was used for the peptide having -OH type carboxyl terminal. The extension of peptide chain by Fmoc-amino acid coupling was performed by N-hydroxybenzo triazole (HOBt)-dicyclohexylcarbodiimide (DCC) method. Particularly, Fmoc-amino acid of amino terminal of each peptide was coupled, and the Fmoc group was removed by using 20% piperidine/NMP (N-methyl pyrrolidone) solution. After washing with NMP and DCM (dichloromethane), the peptide was dried with nitrogen gas. TAF (trifluoroacetic acid)-phenol-thioanisole-H₂O-triisopropylsilane (85: 5: 5: 2.5: 2.5 vol/vol) solution was added thereto. In order to remove protecting group and to separate peptide from resin, the peptide was reacted for 2-3 hours, and it was precipitated by using diethylether.

The crude peptide was purified by using reverse phase(RP)-HPLC column(Delta Pak, C₁₈ 300 Å, 15, 19.0 mm × 30 cm, Waters) in acetonitrile gradient containing 0.05% TFA. Synthesized peptide was hydrolyzed with 6 N-HCl at 110°C, and the residues were vacuum concentrated. And then, its amino acid composition was analyzed with amino acid analyzer (Hitachi 8500 A) after dissolving in 0.02 N-HCl. As a result, the purity of the peptide was over 95%, and the molecular weight obtained by MALDI mass spectrometry (Hill, et al., *Rapid Commun. Mass Spectrometry*, 1991, 5, 395) was the same as the molecular weight obtained by calculation of amino acids. Therefore, it was confirmed that the peptide having correct amino acid sequence was synthesized.

Experimental example 1: Anti-bacterial activity of the peptides

<1-1> Measurement of MIC

In order to measure the anti-bacterial activity of the peptide synthesized in Example 1, minimum inhibitory concentration (MIC) of the peptide was measured.

The present inventors used *Bacillus subtilis* (KCTC 1918) and *Stapilococcus epidermidis* (KCTC 1917) as Gram-

positive bacteria, and *Pseudomonas aeruginosa* (KCTC 1637) and *Salmonella typhimurium* (KCTC 1926) as Gram-negative bacteria for this experiment. All the bacteria used in this experiment were given by Korea Research Institute of Bioscience and Biotechnology (KRIBB). Each bacteria strain was cultured in LB medium (1% bacto-trypton, 0.5% bacto yeast extract, 1% sodium chloride) to the mid-log phase, and diluted with 1% bacto-peptone medium at the concentration of 1×10^4 cells/100 μ l. The diluted bacteria were loaded into micro-titrate plate. Antibiotic peptide synthesized in Example 1 and CA-MA peptide (as a comparative group) were half-fold diluted consecutively from 25 μ M/well, and added into the plate for 6-hour culture at 37°C. Finally, MIC of each strain was determined by observing OD₆₂₀ with micro-titrate plate reader. The results were described in Table 1.

<Table 1>

Anti-bacterial activity of peptides against Gram-positive and Gram-negative bacteria

Peptide	MIC (μ M)			
	Gram-positive		Gram-negative	
	<i>B. subtilis</i>	<i>S. epidermidis</i>	<i>P. aeruginosa</i>	<i>S. typhimurium</i>
CA-MA	3.12	3.12	1.56	0.19
Synthetic peptide (SEQ .ID.NO:2)	0.78	1.56	0.78	0.097

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<1-2> Visualization of anti-bacterial activity

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bacteria.

From the above results, it was confirmed that the synthetic peptide of the present invention represented
5 by the SEQ. ID. NO: 2 had superior antibiotic activity to that of CA-MA peptide.

<1-3> anti-bacterial activity observation with SEM

Anti-bacterial activity of the synthetic peptide
10 of the present invention was observed with SEM (scanning electron microscopy). *Bacillus subtilis* (Gram-positive) and *Pseudomonas aeruginosa* (Gram-negative) cells were cultured in LB medium (1% bacto trypton, 0.5% bacto yeast extract, 1% sodium chloride)
15 to mid-log phase, and the cells were diluted with 10 mM of Na-phosphate buffer containing 100 mM of NaCl) at the concentration of 10^8 cells/ml. Synthetic peptide of the present invention and CA-MA peptide (as a comparative group) were added into the diluted cell
20 culture medium (final conc. 0.78 μ M in *B. subtilis* culture, 1.56 μ M in *P. aeruginosa* culture), followed by further culturing for 30 minutes at 37°C. 0.2 M Na-phosphate buffer containing 5% glutaraldehyde was added into the medium, and the cells were fixed for 2 hours
25 at 4°C. The cells were filtered with isopore filters (0.2 μ m pore size, Millipore, Bedford, MA, USA), and washed with 0.1 M Na-cacodylate buffer (pH 7.4). The

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<1-4> Measurement of membrane dynamic condition

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7.4) was added until OD₄₅₀ reached to 0.25, and DPH (1,6-diphenyl-1,3,5-hexatriene) dissolved in tetrahydrofuran was added (final conc. 10^{-4} M), followed by further culturing for 45 minutes at 37°C. 5 Steady-state fluorescence anisotropy was determined by measuring the strength of fluorescence with spectrofluorometer (HITACHI F-3010, Tokyo, Japan) at 330 nm and 450 nm.

As a result, when *Bacillus subtilis* and 10 *Pseudomonas aeruginosa* were treated with the synthetic peptide of the present invention represented by the SEQ. ID. NO: 2, DPH-labeled fluorescent materials were intercalated 15-20% lower position of membrane comparing to when the cells were treated with CA-MA 15 peptide (Fig. 5).

<Experimental Example 2> Anti-fungal activity of synthetic peptide

<2-1> MTT assay

20 In order to measure the anti-fungal activity of the synthetic peptide of the present invention, the present inventors performed MTT assay with *Candida albicans* (TIMM 1768) and *Tricosporon beigellii* (KCTC 7707). Particularly, PDB medium(20% potato infusion 25 frum, 2% bacto dextrose) containing various fungi was loaded into the wells(100 μ l/well) of 96-well plate. Antibiotic peptides of the present invention and CA-MA

peptide (as a comparative group) were half-fold diluted consecutively, and added into the plate for further culturing. 10 μ l of MTT solution(3-[4,5-dimethyl-2-thiazolyl-2-thiazolyl]-2,5-diphenyl-2H-tetrazolium bromide, 5 mg/ml) was added into each well followed by further culturing for 5-6 hours. Formazan produced by mitochondria enzymes of living cells was dissolved in 100 μ l of 0.04 N HCl-isopropanol. Finally, OD₅₇₀ was measured by using ELISA reader to determine the degree of MIC. The result was described in Table 2.

<Table 2>

Anti-fungal activity of peptides

Peptide	MIC (μ M)	
	<i>C. albicans</i>	<i>T. beigelii</i>
CA-MA	12.5	6.25
Synthetic peptide	6.25	3.25

As a result, it was confirmed that the anti-fungal activity of the synthetic peptide of the present invention represented by the SEQ. ID. NO: 2 was about 2 times higher than that of CA-MA peptide.

<Experimental Example 3> Anticancer activity of antibiotic peptide

In order to measure the anticancer activity of the synthetic peptide of the present invention, the present

inventors performed MTT assay with Calu-6 (a human lung cancer cell line), SNU 601(a human stomach cancer cell line) and Jurkat (a T-cell lymphoma cell line) cells. Firstly, 90 μl of each cell line(2×10^5 cells/ml) was loaded into each well of 96-well plate. At this time, only medium contained wells were used as a control. After shaking well, the cells were cultured in CO_2 incubator for 3 days. Formazan produced by mitochondria enzymes of living cells was dissolved in 100 μl of 0.04 N HCl-isopropanol, and finally, OD_{540} was measured by using ELISA reader. The anti-cancer activity of antibiotic peptide of the present invention was represented by a percentage (OD of synthetic peptide treated well/ OD of control $\times 100$).

As shown in Fig. 6, it was confirmed that the anti-cancer activity of the synthetic peptide of the present invention was higher than that of CA-MA peptide in all cell lines. To the concentration of 1 μM , synthetic peptide of the present invention did not showed anti-cancer activity. However, as concentration increases, the rapidly growing anticancer activity was detected. For example, strong anticancer activity which made complete restrain of cancer cell growth was observed with over 10 μM concentration.

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<Experimental example 4> Cytotoxicity of synthetic peptide

In order to confirm if the synthetic peptide of the present invention showed cytotoxicity, hemolysis capacity of the synthetic peptide was investigated.

Human red blood corpuscles were diluted with PBS (pH 7.0) to the concentration of 8%, and loaded into each wells of 96-well plate. Synthetic peptide of the present invention was half-fold diluted consecutively from 12.5 μ M/well, followed by reacting with the red blood corpuscles for 1 hour at 37°C. After centrifugation, OD₄₁₄ was measured to determine the amount of hemoglobin in the supernatant. At this time, CA-MA peptide was used as a comparative group and melittin was used as a positive control. In order to investigate the level of hemolysis, 1% triton X-100 was added, and then OD was measured. Hemolysis capacity of triton X-100 was regarded as 100%, with which hemolysis capacity of the synthetic peptide was compared and calculated according to the below <Mathematical Formula 1>.

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<Mathematical Formula 1>

$$\% \text{ hemolysis} = (\text{OD A} - \text{OD B} / \text{OD C} - \text{OD B}) \times 100$$

In the above <Mathematical Formula 1>,

25 OD A = OD₄₁₄ of peptide solution,

OD B = OD₄₁₄ of PBS,

OB C = OB₄₁₄ of 1% triton X-100.

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INDUSTRIAL APPLICABILITY

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Those skilled in the art will appreciate that the conceptions and specific embodiments disclosed in the foregoing description may be readily utilized as a

basis for modifying or designing other embodiments for
 carrying out the same purposes of the present invention.
 Those skilled in the art will also appreciate that such
 equivalent embodiments do not depart from the spirit
 5 and scope of the invention as set forth in the appended
 claims.